

Phytoconstituents profiling of Selaginella willdenowii (Desv.) Baker and Pharmacological Potential

by Susilo Wardhani

Submission date: 12-Jan-2024 02:34PM (UTC+0700)

Submission ID: 2269817238

File name: 60_RJPT_16_11_2023.pdf (264.18K)

Word count: 6157

Character count: 33264

RESEARCH ARTICLE

Hippobroma longiflora (L.) G. Don: Comparative Phytochemical Screening and Potential Activities of Flower and Leaf by GC-MS

22

Nabilla Sinta Dewi, Susilo Susilo*

 Department of Biology Education, Faculty of Teacher Training and Education,
 Universitas Muhammadiyah Prof. DR. HAMKA, East Jakarta, Indonesia 13830.

*Corresponding Author E-mail: susilo@uhamka.ac.id

29 STRACT:

Hippobroma longiflora (L.) G. Don or "kitolod" (Indonesia) is widely known as a traditional food medicine. However, the bioactive constituents of this plant are still unclear. Therefore, we investigated the bioactive compounds of *H. longiflora* (leaves and flowers) with Gas Chromatography-Mass Spectrometry (GC-MS) and further evaluated their potential biological activity. Ethanol extracts from the flowers and leaves of *H. longiflora* are targeted for phytochemical profiling of this plant. As a result, 16 metabolite compounds on leaves and 21 on flowers were successfully detected. A total of seven compounds, namely 2, 6, 10, 14, 18, 22-. Tetracosahexaene, 2, 6, 10, 15, 19, 23-hexamethyl-, (all-E)-, 2,6,10-Trimethyl, 14-Ethylene-14-Pentadecne, Stigmast-5-EN-3- Stigmasterol, and Vitamin E dominate in both parts of this plant. The results of phytochemical analysis at the National Institute of Standard and Technique (NIST), *H. longiflora* can be an essential source of anti-tumor, anti-cancer, antibacterial, and antioxidant drugs.

KEYWORDS: Bioactive constituents, *Hippobroma longiflora* (L.) G. Don, GC-MS, kitolod, phytochemicals.

INTRODUCTION:

Medicinal plants contain traditional bioactive compounds beneficial to humans in curing various diseases¹. Medicinal plants have been used since ancient times² as a tonic, food, or to increase immunity and body strength²⁻⁵. It has the characteristic of a thin and low rosette⁶⁻⁷. Green leaves with toothed linear and curved growth and white flowers with elongated tubular⁸. The plant belongs to Campanulaceae and spreads in tropical, temperate, and cold regions^{9,10}.

The flower and leaf parts of *H. longiflora* (L.) G. Don is commonly used in medicine and food. The ethanol extract of *H. longiflora* (L.) G. Don leaves alkaloids, flavonoids, polyphenols, monoterpenoids, sesquiterpenoids, quinine, steroids, coumarins, tannins, and saponins¹¹⁻¹³. The compound (-)-Lobeline is known that have been isolated from *Lobelia inflata*, extensively studied its structure and biological activity¹⁴, and is thought to have many functional similarities with the genus *Hippobroma*¹⁵.

Bioactive compounds in *H. longiflora* (L.) G. Don flower extract is a group of fatty acids and their derivatives¹⁶.

Pharmacologically, the flavonoid content of *H. longiflora* (L.) G. Don can be used as a natural sunscreen and inhibit the development of cataracts¹⁷⁻¹⁹. Other uses of *H. longiflora* (L.) G. Don is also an antioxidant, anti-cancer, anti-inflammatory, antibacterial, anti-diabetic, antimalarial, anti-tumor, antimicrobial, antifungal, anti-insecticide, and antiseptic²⁰⁻²². Other studies prove that the bioactive constituents of *H. longiflora* (L.) G. Don can be used as a folk remedy for bronchitis, laryngitis, asthma, wounds, eye medication, anti-neoplastic, hemostasis, and analgesic^{23,24}.

Recent reports state that a 75% concentration of ethanol extract from *H. longiflora* (L.) G. Don leaves effectively inhibit the growth of *Staphylococcus aureus* and *Salmonella typhi* with robust criteria^{25,26}. Other studies also mentioned that ethanol extract from *H. longiflora* (L.) G. Don leaves inhibits cornea neovascularization after seven-day administration to the eyes that have experienced chemical trauma due to the pesticide Rotraz© 200EC²⁷. *H. longiflora* (L.) G. Don is documented in Indonesian pharmacology as one of the

Received on 26.01.2023 Modified on 18.05.2023
 Accepted on 14.09.2023 © RJPT All right reserved
 Research J. Pharm. and Tech 2023; 16(11):5347-5353.
 DOI: 10.52711/0974-360X.2023.00866

traditional herbs (JAMU) by the Indonesian population. They are traditionally extracted by the natives to treat human diseases. However, the complete constituents of this plant are still unclear. Hence, this study aims to compare the phytochemical constituents of *H. longiflora* (L.) G. Don (leaves and flowers) with GC-MS analysis and further evaluate the potential for biological activity.

34

MATERIALS AND METHODS:

Plant Materials:

Fresh plants of *H. longiflora* (L.) G. Don were collected from farmers' gardens in Mustika Jaya District, Bekasi, West Java, Indonesia (6°17'31"S, 107°1'46"E) in August 2022. This plant is authenticated at ELSA, Indonesia Botani Identification Services, and specimens are deposited at the university with voucher number 1961537. The sample was washed and stored at 20°C until subsequent analysis.

Sample Extraction Preparation:

Each part was weighed by 100g and oven dried at 33.5°C for 45 minutes following procedure²⁸. Dried samples were mashed with a grinder and stored at -18°C in low-density polyethylene bags²⁹. For three days, the stock concentration of 20mg/mL extract was prepared by soaking a powder sample of 20g of powder (40 mesh) in Pro Analyst²⁴ ethanol (99.8%). The solution mixture is stirred and placed in a 40°C waterbath and shaken at 250rpm for 1hour³⁰. The mixture was centrifuged (2000rpm) for 10min, followed by filtration. The residue was immersed in a solvent and evaporated at 65°C using Rotary Evaporator. The extraction results are stored inside the fume hood to evaporate the excess solvent further until it produces a stickier and more concentrated extract. The extract was held at 4°C until further analysis³¹.

GC-MS Condition:

Gas Chromatography-Mass Spectrometry (GC-MS) was used to decompose the phytoconstituents of target plants³². A combination of a gas chromatograph 7890 A (Agilent 19091-433HP, USA) and a mass spectrophotometer equipped with an HP-5 MS molten silica column connected to a 5675C-MS-D and a Three Axis detector was implemented³³. The capillary column of GC-MS HP-5MS (30m 0.25mm ID 0.25m) consists of 5% diphenyl and 95% Dimethylpolysiloxane²⁸. The ethanol extract³¹ was filtered using a 5µL filter in split mode (8:1). Helium gas was used as the carrier at a rate of 1.2mL/min at an injector temperature of 250°C. Then the analyte is separated into a silica capillary column. The oven program is set up as follows: an initial

temperature of 80°C which is directly¹ raised by 3°C/min to 150°C and held for 1 minute, then increased by 20°C/min to 280°C³⁵ and held for 26minutes²⁸. Ionization energy (70eV) was used to determine the mass spectrum.

Data Analysis:

The Agilent Mass Hunter software analyzes bioactive compounds by comparing mass fragments and the standard mass spectrum. Biological activity data were analyzed using the PubChem library database, Chemistry WebBook, and FOODB from the Metabolomics Innovation Center. This library was also used in Guang-Mei Tang's research in 2022³⁴.

RESULTS AND DISCUSSION:

This study shows that the GC-MS method is a powerful combination of phytoconstituents composition analysis^{5,35-37}. The components can be separated effectively by GC and MS completing thorough component identification^{12,38-41}. We agree that the GC-MS method provides an excellent opportunity to explore phytoconstituents as has been done by many previous researchers^{26,28,50,42-49}. This study used the GC-MS to determine the phytoconstituents of *H. longiflora* (L.) G. Don leaf and flower extracts. The¹ GC-MS chromatogram of *H. longiflora* (L.) G. Don recorded a total of 37 peaks corresponding to phytochemical compounds recognized by attributing Retention Time (RT) and Concentration (%). As a result, as many as 16 unique metabolite compounds were present in the leaf and 21 in the flower of *H. longiflora*. The results of the leaf chromatogram of *H. longiflora* (L.) G. Don showed the highest RT value in Squalen from the Prenol lipids group (Figure 1).

9

-3,7,11,Trimethyl-dodeca-2,4,6,10-Tetraenal- is a unique compound that is only present on leaves with two different time retention. This lipid prenol derivative is a product of lycopene oxidation⁵¹ in biological tissues and has been studied to have a role as an anti-depressant. Another unique compound² detected that also has two retention times is 12-Methoxy-18-Nor-5.Beta-Podocarpa-8,11,13-Trien-3.Alpha. O.L., a derivative of the Hydrophenanthrenes group, has yet to be discovered in terms of its benefits. Interpretation of the mass spectral chromatogram of *H. longiflora* (L.) G. Don compounds is presented in **Table 1**.

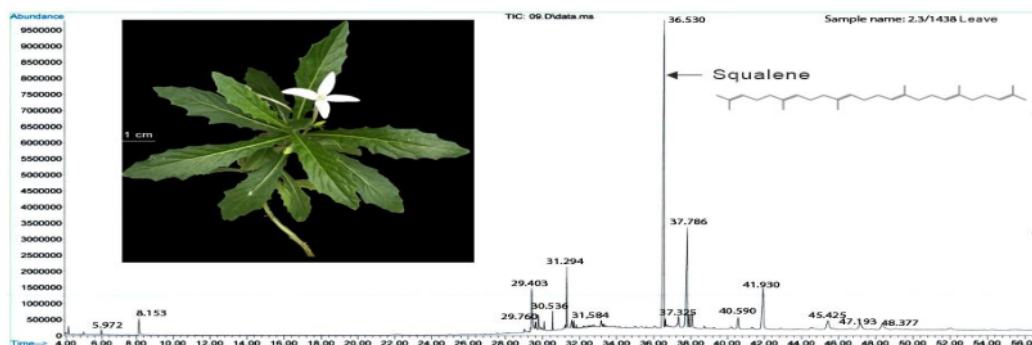


Figure 1: Chromatogram of *H. longiflora* (L.) G. Don leaf

Table 1: List of compounds leaf and flower of *H. longiflora* (L.) G. Don

M.F.	Compound	M.W.	Classification	Leaf		Flower	
				R.T.	% of Area	R.T.	% of Area
C ₇ H ₆ O 27	Benzaldehyde	106.12	Benzene and substituted derivatives	-	-	5.866	1.2
C ₃₀ H ₅₀ O	(2E)-3,7,11,15--Tetramethyl-2-Hexadecene--1-OL	296.5	Prenol lipids	31.296	4.17	-	-
C ₂₆ H ₄₃ N ₃ O ₄	(2E)-9-Octadecenoic Acid	489.7	Fatty acyls	31.585	1.17	-	-
C ₁₅ H ₁₆ O ₂ S	12-Methoxy--18-Nor-5.Beta. --Podocarpa-8,11,13-Trien-3.Alpha.-OL	260.4	Hydrophenanthrenes	-	-	37.370	1.89
C ₆ H ₅ COCH ₃	1-Phenylethanone	120.15	Organooxygen compounds	8.155	1.65	-	-
C ₁₈ H ₃₈	2,6,10-Trimethyl-, -14-Ethylene-14-Pentadecne	278.5	terpenoid	29.399	5.64	29.399	1.12
C ₁₄ H ₁₆ O ₂ S	2,4',6-Trimethyldiphenylsulfone	260.4	Not Found	37.784	14.81	-	-
C ₃₀ H ₅₀	2,6,10,14,18,22--Tetracosahexaen, 2,6,10,15,19,23--hexamethyl-(all-E)-	410.7	Prenol lipids	36.529	39.01	36.515	7.54
C ₆ H ₁₀ N ₂ O ₄	2-[4-(1-Methylethoxy)bnzl]-4,6-pyrimidinediol	210.19	Benzene and substituted derivatives	37.895	1.5	-	-
C ₁₈ H ₁₇ O ₂	2,3-dihydroxy-2-Phenyl-1-Acenapthenon	260.29	Naphthalenes	37.322	1.85	-	-
C ₁₅ H ₂₂ O	3,7,11,-Trimethyl-dodeca-2,4,6,10,7-raenal	218.33	Prenol lipids	29.613	1.24	-	-
C ₃₀ H ₄₈ O	4,4,6a,6b,8a,11,12,14b,Octamethyl-1,4,4a,5,6,6a,6b,7,8,8a,9,10,11,12,12a,14,1,5,14b-Octadecydro-2H-Picen-3-one	424.7	Prenol lipids	-	-	49.272	2.27
C ₁₉ H ₂₂ O ₆	6,7,8,9-Tetrahydro--3-Methoxymethyl-5,9-Dimethyl-8-Hydroxy-Napho [1,2-B]ran	346.4	Naphthofurans	38.067	2.34	-	-
C ₁₇ H ₁₃ N ₂ O	6H-Pyrido- [4,3-B] Carbazole-5-oxaldehyde, 11-Methyl-	260.29	Carbazoles	-	-	38.032	41.3
C ₁₀ H ₁₈	8-Methyl--7-Thiabicyclo [4.2.1] nonan	138.25	Not Found	-	-	43.018	1.33
C ₃₉ H ₇₀ O ₆	9,12-Octadecadienoic Acid	635.0	Fatty Acyls	-	-	36.143	1.08
C ₁₀ H ₂₁ NO	Acetamine, N-(1,1,3,3-tetramethyl butyl)-	171.28	Indoles and derivatives	-	-	38.825	38.825
C ₈ H ₈ O	Acetophenone	120.15	Organooxygen compounds	-	-	8.127	5.04
C ₃₀ H ₅₀ O	Alpha Amyrin	426.7	Prenol lipids	48.375	2.23	-	-
C ₃₁ H ₆₄	Hentriacontane	436.8	Saturated hydrocarbons	-	-	40.701	3.87
C ₃₀ H ₄₂	Hexadecane, 2,6,10,14-Tetramethyl-	282.5	Diterpenes	-	-	42.873	2.45
C ₂₀ H ₄₂	Icosane	282.5	Saturated hydrocarbons	-	-	35.040	1.85
C ₂₉ H ₆₀	Nonacosane	408.8	Saturated hydrocarbons	-	-	37.288	1.89
C ₁₉ H ₄₀	Nonadecane	268.5	Saturated hydrocarbons	-	-	32.261	1.76
C ₂₈ H ₅₈	Octacosane	394.8	Saturated hydrocarbons	40.591	1.93	-	-
C ₂₉ H ₅₀ O	Stigmast-5-EN-3-OL-	414.7	Steroids and steroid derivatives	47.196	2.23	47.548	3.54
C ₂₉ H ₅₀ O ₂	Vitamin E	430.7	Prenol lipids	41.928	8.09	51.078	1.44
C ₂₉ H ₄₈ O	Stigmasterol	412.7	Steroids and steroid derivatives	45.424	2.55	42.038	4.77
C ₁₆ H ₃₀ O	Z,Z-11,13-Hexadecadien-1-ol	238.41	Fatty acid derivative	-	-	32.516	1.57

Table 2: Equation of compounds in extracts of leaves and flowers of *H. longiflora* (L.) G. Don

Compounds	Leaf		Flower	
	R.T.	% of Area	R.T.	% of Area
2,6,10,14,18,22-Tetracosahexaene, 2,6,10,15,19,23-hexamethyl-, (all-E)-	36.529	39.01	36.515	7.54
2,6,10-Trimethyl, 14-Ethylene-14-Pentadecne	29.399	5.64	29.399	1.12
3,7,11,Trimethyl-dodeca-2,4,6,10-Tetraenal	29.613	1.24	-	-
12-Methoxy-18-Nor-5.Beta.-Podocarpa-8,11, -13-Trien-3.Alpha.OL	29.758	1.82	-	-
Stigmast-5-EN-3-OL	-	-	37.370	1.89
	-	-	38.260	2.62
Stigmast-5-EN-3-OL	47.196	2.23	47.548	3.54
	-	-	51.078	1.44
Stigmast-5-EN-3-OL	45.424	2.55	45.672	2.64
Vitamin E	41.928	8.09	42.038	4.77

The main compounds in the leaf extract found with a high percent peak area are 2,6,10,14,18,22-Tetracosahexaene, 2,6,10,15,19,23-hexamethyl-, (all-E)- or Squalene compounds (39.01%), 2,4',6-Trimethyl-diphenylsone (14.81%), Vitamin E (8.09%), and 2,6,10-Trimethyl, 14-Ethylene-14-Pentadecne (5.64%), (2E)-3,7,11,15-Tetramethyl-2-Hexadecene-1-OL- (4.17%). There are several compounds with almost the same peak area, namely Stigmast-5-EN-3-OL (peak area: 2.55%), 6,7,8,9-Tetrahydro-3-Methoxymethyl-5,9-Dimethyl-8-Hydroxy-Naphtho-[1,2-B] Furan (2.34%) and two compounds with the same peak Stigmast-5-EN-3-OL and Alpha Amyrin (2.23%).

The compound 6H-Pyrido [4,3-B] Carbazole-5-Carboxaldehyde, 11-Methyl- belongs to the class Carbazoles and is the main phytoconstituent in the flower *H. longiflora* (L.) G. Don, indicating the presence of a dominant influence of function on the flower. Other compounds with a reasonably high peak are Vitamin E (4.77%) and hentriacontane (3.87%). The rest of the compound was found with almost the same peak area. 9,12-Octadecadienoic acid (1.08%) was found to be the compound with the lowest biological activity in the extracted compound in *H. longiflora* (L.) G. Don flowers. The nonane 8-Methyl-7-Thiabicyclo [4.2.1] compound is a new compound that cannot be detected in the database.

From a total of 37 compounds isolated from *H. longiflora* (L.) G. Don leaf and flower extracts, there are 7 compound variant equations with different percentages and retention times (Table 2). Compounds 2,6,10,14,18,22-Tetracosahexaene, 2,6,10,15,19,23-hexamethyl-, (all-E)-, 2,6,10-Trimethyl, 14-Ethylene-14-Pentadecne, Stigmast-5-EN-3-OL, Stigmast-5-EN-3-OL, and Vitamin E are bioactive compounds found in flowers as well as leaves. The retention time of compounds 2,6,10,14,18,22-Tetracosahexaene, 2,6,10,15,19,23-hexamethyl-, (all-E)- on the leaves are not much more significant when compared to the same compound in the flower part, and this suggests that the leaves take longer

to come out. The compound also has a high percentage of areas in the leaf part. As for the compound 2,6,10-Trimethyl, 14-Ethylene-14-Pentadecne has the same retention time on the leaf part of the flower. The repeated retention time belongs to the compound Stigmast-5-EN-3-OL; in the leaves, this compound has only one-time retention, but in flowers, it has two-time retention. The bioactive constituents of stigmast-5-EN-3-OL and Vitamin E indicate a stable retention time in the leaves and flowers of *H. longiflora* (L.) G. Don. The compound 3,7,11,Trimethyl-dodeca-2,4,6,10-Tetraenal was found only on the leaves alone but had a different time interval to exit the column and reach the detector, while 12-Methoxy-18-Nor-5.Beta.-Podocarpa-8,11,13-Trien-3.Alpha. O.L. is only found in the flower section and has a different retention time.

Various potentials are found in the content of leaf extracts and flowers of *H. longiflora* (L.) G. Don (Table 3). Squalene or by other names 2,6,10,14,18,22-Tetracosahexaen, -2,6,10,15,19,23-hexamethyl-, (all-E)- (peak area: 39,01%) is a natural triterpene hydrocarbon. It is an essential intermediary in intensifying many bioactive secondary metabolites⁵². This compound has various benefits for human health, such as anti-cancer effects, cardiovascular disease, and boosting the immune system⁵³. In addition, it also has antibacterial properties⁵⁴.

In *H. longiflora* (L.) G. Don flowers, there are bioactive compounds, namely 6H-Pyrido [4,3-B] Carbazole-5-Carboxaldehyde, 11-Methyl- (41.3%) or with the compound name ellipticine. This bioactive constituent is a pyridocarbazole type plant alkaloid that exhibits cytotoxic activity against tumor cells. It is known that ellipticine has exciting photophysical properties in different solvents⁵⁵. It has pharmacological potential as an anti-tumor and anti-neoplastic⁵⁶. Anti-neoplastic agents are drugs that prevent, kill, or inhibit the growth and spread of cancer cells. Most anti-neoplastic agents act on molecular targets important for cell division⁵⁷.

Then it was found that there were eight compounds in flowers and leaves whose biological activity was unknown. Compound -12-Methoxy-18-10-5-Beta-Podocarpa-8,11,13-Trien-3. Alpha.-OL, 8-Methyl-7-Thiabicyclo- [4.2.1] nonan, 9,12-Octadecadienoic Acid, Hexadecane, 2,6,10,14-Tetramethyl-, Nonadecane on flowers has not found its biological activity. Other

compounds in the leaves that until now are still unknown in biological activity are 2-[4-(-1 Methyleneoxy) bnzt]-4,6-pyrimidinol, 2-Hydroxy-2-Phenyl-1-Acenaphthenon, -6,7,8,9-Tetrahydro-3-Methoxymethyl-5, -9-Dimethyl-8-Hydroxy-Naphtho [1,2-B] Furan.

Table 3: Biological activities of *H. longiflora* (L.) G. Don

Compounds	Part	Biological Activity
11 aldehyde	Flower	Flavoring agents ⁵⁸ and irritants ⁵⁹
(2E)-3,7,11,15-Tetramethyl-2-Hexadecene-1-OL	Leaf	Antioxidant ⁶⁰ , anti-infective and antiparasitic ⁶¹
2)-9-Octadecenoic Acid	Leaf	Antifungal ⁶²
12-Methoxy-18-Nor-5.Beta.-Podocarpa-8,11,13-Trien-3. Alpha.-OL	Flower	Not Found
12-methyleneone	Leaf	Anti-diabetic and anti-adipogenic ⁶³
2,6,10-Trimethyl, 14-Ethylene-14-Pentadecne	Leaf and Flower	Antioxidant, Antiproliferative, and Anti-cancer ⁶⁴
2,4',6-Trimethyl-25-enylsulfone	Leaf	Antibacterial and anti-cancer ⁶⁵
2,6,10,14,18,22-Tetracosahexaen, 2,6,10,15,19,23-hexamethyl-(all-E)-	Leaf and Flower	Antibacterial ⁶⁶ and insecticide ⁶⁶
2-[4-(-1 Methyleneoxy) bnzt]-4,6-pyrimidinediol	Leaf	Not Found
9-Hydroxy-2-Phenyl-1-Acenaphthenon	Leaf	Not Found
8-11, Trimethyl-dodeca-2,4,6,10-tetraenol	Leaf	Anti-depressants ⁶⁷
4,4,6a,6b,8a,11,12,14b-Octamethyl-1,4,4a,5,6,6a,6b,7,8,8a,9,10,11,12,12a,14,14a,14b-Octahydro-2H-Pyridin-3-one-	Flower	Anti-cancer ⁶⁸
6,7,8,9-Tetrahydro-3-Methoxymethyl-5,9-Dimethyl-8-Hydroxy-3-phtho [1,2-B] Furan	Leaf	Not Found
10-Pyrido [4,3-B] - Carbazole-5-Carboxaldehyde, 11-Methyl-8-Methyl-7-Thiabicyclo [4.2.1] nonan-	Flower	Antitumor dan antineoplastic ⁶⁹
9,12-Octadecadienoic Acid	Flower	Not Found
Acteamine, N-(1,1,3,3-tetramethylbutyl)-	Flower	Organic synthesis and chemical medicine ⁶⁹
Acetophenone	Flower	Antifungal ⁷⁰ and flavoring agents ⁷¹
Alpha Amyrin	Leaf	Anti-inflammatory, anxiolytic, and anti-depressant ⁷²
Hentriacontane	Flower	Anti-inflammatory, anti-tumor, and Antibacterial ⁷³
Hexadecane, 2,6,10,14-Tetramethyl-	Flower	Not Found
Icosane	Flower	It has thermal properties ⁷⁴
Nonacosane	Flower	The main paraffin of most plant waxes ⁷⁴
Nonadecane	Flower	Not Found
Octacosane	Leaf	Antioxidant ⁷⁵
Stigmast-5-EN-3-OL	Leaf and Flower	Anti-diabetic, cholesterol reduction and anti-cancer ⁷⁶
Stigmastrol	Leaf and Flower	Immune response, increasing the concentration of enzymes ⁷⁷ , and -anti-cancer ⁷⁸
Vitamin E	Leaf and Flower	Chronic disease prevention, antioxidants, and neurological therapy ⁷⁹
ZZ-11,13-Hexadecadien-1-ol	Flower	Medical, pharmaceutical, and cosmetic components ⁸⁰

CONCLUSION:

This study was one of the few reports in which the bioactive compound extracts of *H. longiflora* (L.) G. Don were reported. Various bioactive compounds were found in *H. longiflora* (L.) G. Don. Thirty-seven chemical constituents of *H. longiflora* (L.) G. Don leaf and flower extracts were identified with GC-MS. Squalene, 2,4',6-Trimethyldiphenylsulfone and Vitamin E which predominate in the leaves act as a producer of high amounts of antioxidants and also as a medium for cancer treatment. Ellipticine, Vitamin E, and hentriacontane predominate in flowers with their role as anti-neoplastic agents that can prevent, kill, or inhibit the growth and spread of cancer cells. Squalene and other bioactive compounds highlight the use of drugs

and pharmaceuticals. Based on phytochemicals identified with potential biological activity, this plant can be an essential ingredient in medicine. Its ability as an anti-tumor, anti-cancer, antibacterial, and antioxidant drug from this plant as a natural constituent source can be explored further.

ACKNOWLEDGMENTS:

The author would like to thank the ELSA Botanical Identification Services and Herbarium Bogoriense, National Research and Innovation Agency (BRIN), Indonesia.

COMPETING INTERESTS:

The authors declare that they have no competing interests.

REFERENCES:

- Bhattacharya R, Naitam P. Green Anticancer Drugs-An Review. Res J Pharmacogn Phytochem. 2019; 11(4): 231. doi: 10.5958/0975-4385.2019.00040.2
- Aziz IR, Raharjeng ARP, Susilo S, Nasution J. Ethnobotany of traditional wedding: A comparison of plants used by Bugis, Palembang, Sundanese and Karo ethnic in Indonesia. J Phys Conf Ser. 2019; 1175(1). doi: 10.1088/1742-6596/1175/1/012005
- Bhalla N, Ingle N, Patri S V., Haranath D. Phytochemical analysis of Moringa Oleifera leaves extracts by GC-MS and free radical scavenging potency for industrial applications. Saudi J Biol Sci. 2021; 28(12): 6915-28. doi: 10.1016/j.sjbs.2021.07.075
- Akbar B, Susilo S, Nissa RA, Ritonga RF, Lestari S, Astuti Y, Parwito P. Antifertility Effect of the Ethanol Extract of Centella asiatica L. Urban Against the White Rat (*Rattus norvegicus* L.) in the Early Post-Implantation. J Phys Conf Ser. 2018; 1114(1). doi: 10.1088/1742-6596/1114/1/012002
- Bele AA, Khale A. An approach to a nutraceutical. Res J Pharm Technol. 2013; 6(10): 1161-4.
- Folquitto DG, Swiech JND, Pereira CB, Bobek VB, Halila Possagno GC, Farago PV, Miguel M, Duarte JL, Miguel OG. Biological activity, phytochemistry and traditional uses of genus *Lobelia* (Campanulaceae): A systematic review. Fitoterapia. 2019; 134: 23-38. doi: 10.1016/j.fitote.2018.12.021
- Brantjes NBM. Regulated Pollen Issue In *Isotoma*, Campanulaceae, and Evolution Of Secondary Pollen Presentation. Acta Bot Neerl. 1983 May;32(3):213-22. doi: 10.1111/j.1438-8677.1983.tb01702.x
- Lammers TG. Revision of the Infrageneric Classification of *Lobelia* L. (Campanulaceae: Lobelioideae). Annals of the Missouri Botanical Garden. 2011 Apr;98(1):37-62. doi: 10.3417/2007150
- Pe'a R. Conserving Hawaiian Lobeliads: The Story of *Cyanea tritomantha*, an Endemic Hawaiian Campanulaceae. Imperiled Encycl Conserv. 2022;823-30. doi: 10.1016/b978-0-12-821139-7.00132-x
- Koutsovoulou K, Daws MI, Thanos CA. Campanulaceae: a family with small seeds that require light for germination. Ann Bot. 2014 Jan;113(1):135-43. doi: 10.1093/aob/mct250
- Anjelina SH. Antibacterial Activity of Ethanol Extract of Kitolod (*Hippobromalongiflora*) Leaf Against *Staphylococcus aureus* and *Salmonella typhi*. Asian J Pharm Res Dev. 2020; 8(1):52-4. doi: 10.22270/ajprd.v8i1.660
- Wiranto B, Husnin, Susilo. Diversity of terrestrial ferns (Pteridophytes) in Ciliwung Telaga Warna Puncak Bogor tea estate in West Java. J Phys Conf Ser. 2021; 755(1): 1-6. doi: 10.1088/1755-1315/755/1/012031
- Nurmawati D, Sudiarti D, Hasbiyati H. Identification of Medicinal Plant Potential of Kasiyan Village Puger District. Bioeduscience. 2022;6(1):31-40. doi: 10.22236/j.bes/617819
- Chan ZY, Govindaraju K, Krishnan P, Low YY, Chong KW, Yong KT, Ting KN, Lim KL. Diphenethylpiperidine alkaloids with tracheal smooth muscle relaxation activity from *Hippobroma longiflora* (L.) G. Don. 2019; 30: 93-98. doi: 10.1016/j.phytol.2019.01.035
- Tadege G, Alebachew Y, Hymete A, Tadesse S. Identification of lobetyolin as a major antimalarial constituent of the roots of *Lobelia giberroa* Hemsl. Int J Parasitol Drugs Drug Resist. 2022; 18: 43-51. doi: 10.1016/j.ijpddr.2022.01.002
- Siregar RM. Aktivitas antibakteri ekstrak daun dan bunga kitolod (*laurentia longiflora* (L.) P. term) terhadap beberapa bakteri penyebab konjungtivitas. 2012; <http://repository.ipb.ac.id/handle/123456789/56297>
- Fitriani D. Overview of COVID-19 Infection Manifestation in Neuropsychiatry Aspect. Open Access Indonesian Journal of Medical Reviews. 2021; 1(3): 49-52. doi: 10.37275/oajimr.v1i3.40
- Martiningih NW, Mudiarta IW, Suryanti IAP. Phytochemical screening and antioxidant activity of *hippobroma longiflora* extracts. IOP Conf Ser Mater Sci Eng. 2021; 1115: 012078. doi: 10.1088/1757-899X/1115/1/012078
- Ramses, Fenny Agustina, R. Pramuanggit Panggih Nugroho. Antibacterial Potential of Bidara Laut (*Ximenia americana*) Plant Against *Vibrio alginolyticus* and *V. parahaemolyticus* Bacteria. Bioeduscience. 2021; 5(1): 15-23. doi: 10.22236/j.bes/515091
- Arrosyd M, Mustofa CH, Sutaryono, Rohmah AP. Effect of boiling time on content of the total flavonoid of kitolod (*Isotoma longiflora* (L.) C. Presl.). J Phys Conf Ser. 2021; 1764(1): 012023. doi: 10.1088/1742-6596/1764/1/012023
- Hernawati D, Dwisandi RF, Nuryadin E. Potential of Bioactive Compounds of Arenga Vinegar as Traditional Medicine Through Reverse Docking Techniques. Bioeduscience. 2021; 5(2): 142-7. doi: 10.22236/j.bes/526802
- Karpagam T, Firdous J, Revathy, Priya S, Varalakshmi B, Gomathi S, Geetha S, Muhamad N. Anti-Cancer Activity of A loe Vera Ethanol Leaves Extract against In vitro Cancer Cells. Res J Pharm Technol. 2019; 12(5): 2167. doi: 10.5958/0974-360X.2019.00360.3
- Haryoto H, Hapsari A. Cytotoxicity of ethanol extract, polar, semipolar, and nonpolar herb kitolod (*Isotoma longiflora* (L.) C. Presl.) Cells on MCF-7 Cells. 2019; 603-9.
- Tanaja GI, Isbandiati E, Wattimena I. Antibacterial activity of ethanolic extract of kitolod (*Hippobroma longiflora*) leaves against streptococcus pyogenes. J Widya Med Jr. 2022; 4(1):56-60.
- Pawar RS, Dimri M, Maitihani A, Kush L. The application of marine natural products (MNPS) in anti-covid-19 therapeutics. Asian J Pharm Res Dev. 2020; 8(6):77-80. doi: 10.22270/ajprd.v8i6.809
- Bhavani R, Bhuvanewari E, Rajeshkumar S. Antibacterial and antioxidant activity of ethanolic extract of ceiba pentandra leaves and its phytochemicals analysis using GC-MS. Res J Pharm Technol. 2016; 9(11): 1922-6. doi: 10.5958/0974-360X.2016.00393.0
- Arsyad HM, Komariah C, Hasan M. The effect of isotoma longiflora leaves extract to the Cornea Neovascularization of Wistar Rats chemical trauma model. J Agromedicine Med Sci. 2020; 6(2): 92. doi: 10.19184/ams.v6i2.6853
- Nabila N, Susilo S. A Comparative metabolite analysis of *Pandanus amaryllifolius* leaves from different growth stages using GC-MS and Their Biological. Eur Chem Bull. 2022; 11(12): 22-38. doi: 10.31838/ecb/2022.11.12.003
- Dauber C, Carreras T, González L, Gámbaro A, Valdés A, Ibanez E, Vieitez I. Characterization and incorporation of extracts from olive leaves obtained through maceration and supercritical extraction in Canola oil: Oxidative stability evaluation. LWT. 2022; 160: 113274. doi: 10.1016/j.lwt.2022.113274
- Ola SS, Catia G, Marzia I, Francesco VF, Afolabi AA, Mulinacci N. HPLC/DAD/MS characterisation and analysis of flavonoids and cinnamoid derivatives in four Nigerian green-leafy vegetables. Food Chem. 2009;115(4):1568-74. doi: 10.1016/j.foodchem.2009.02.013
- Goh MPY, Kamaluddin AF, Tan TJL, Yasin H, Taha H, Juma J, Ahmad N. An evaluation of the phytochemical composition, antioxidant and cytotoxicity of the leaves of *Litsea elliptica* Blume - An ethnomedicinal plant from Brunei Darussalam. Saudi J Biol Sci. 2022;29(1):304-17. doi: 10.1016/j.sjbs.2021.08.097
- Akar Z. Chemical compositions by using LC-MS/MS and GC-MS and antioxidant activities of methanolic extracts from leaf and flower parts of *Scabiosa columbaria* subsp. *columbaria* var. *columbaria* L. Saudi J Biol Sci. 2021; 28(11): 6639-44. doi: 10.1016/j.sjbs.2021.07.039
- Olivia NU, Goodness UC, Obinna OM. Phytochemical profiling and GC-MS analysis of aqueous methanol fraction of *Hibiscus asper* leaves. Futur J Pharm Sci. 2021;7(1). doi:10.1186/s43094-021-00208-4
- Tang GM, Shi YT, Gao W, Li MN, Li P, Yang H. Comparative analysis of volatile constituents in root tuber and rhizome of *Curcuma longa* L. using fingerprints and chemometrics approaches on Gas Chromatography-Mass Spectrometry. Molecules. 2022; 27(10):3196. doi: 10.3390/molecules27103196
- Pawar AR, Vikhe DN, Jadhav RS. Recent Advances in Extraction Techniques of Herbs - A Review. Asian J Res Pharm Sci. 2020;10(4):287-92. doi: 10.5958/2231-5659.2020.00050.8
- Thakur P, Thakur U, Kaushal P, Ankalgil AD, Kumar P, Kapoor A, Ashawat MS. A Review on GC-MS Hyphenated Technique. Asian J Pharm Anal. 2021; 11(4): 285-92. doi: 10.52711/2231-5675.2021.00049
- Phale MD, Korgaonkar D. Current Advance Analytical Techniques: A Review. Asian J Pharm Anal. 2009; 2(3): 235-8.
- Aravind R, Bindu AR, Bindu K, Alexeyeva V. GC-MS analysis of the bark essential oil of *cinnamomum malabattrum* (burman. f) blume. Res J Pharm Technol. 2014; 7(7): 754-9.
- Susilo S, Setyaningsih M. Analysis of genetic diversity and genome relationships of four eggplant species (*Solanum melongena* L.) using RAPD markers. J Phys Conf Ser. 2018; 948(1):0-6. doi: 10.1088/1742-6596/948/1/012017
- Susilo S, Meityani. Genetic variation of three *brugiera* species from Karimunjawa Islands detected by using RAPD molecular markers. Asian J Plant Sci. 2018; 17(4): 198-203. doi: 10.3923/ajps.2018.198.203
- Risnawati R, Meityani, Susilo S. The effect of adding Kepok Banana peels (*Musa paradisica*) to powder media on the growth of white oyster mushrooms (*Pleurotus ostreatus*). J Phys Conf Ser. 2021; 755(1). doi: 10.1088/1755-1315/755/1/012066
- Reddy MY, Ramesh V, Reddy CK, Venugopal N, Saravanana G, Suryanarayana MV. The Quantitative Determination of Process Related Genotoxic Impurities in Esomeprazole Magnesium by GC-MS. Asian J Pharm Anal. 2011; 4(6): 898-901.
- Priya S, Nethaji S, Sindhuja B. GC-MS analysis of some bioactive constituents of diospyros Virginiana. Res J Pharm Technol. 2014; 7(4): 429-32.
- Rajabudeen E, Ganthi A, Subramanian M. GC-MS Analysis of the Methanol Extract of *Tephrosia villosa* (L.) Pers. Asian J Res Chem. 2012; 5(11): 1331-4.
- Zahi MR, Liang H, Khan A, Yuan Q. Identification of Essential Oil Components in Chinese Endemic Plant *Achnatherum inebrians*. Asian J Res

- Chem. 2014; 7(6): 576–9. doi: 10.5958/0974-4150.2018.00150.5
46. Jose BE, Selvam PP. Identification of Phytochemical Constituents in the Leaf Extracts of Azima tetracantha Lam using Gas Chromatography-Mass Spectrometry (GC-MS) analysis and Antioxidant Activity. *Asian J Res Chem.* 2018; 11(6): 857. doi: 10.5958/0974-4150.2018.00150.5
 47. Krishnaveni M, Krishna Kumari G, Ragina Banu C, Kalaivani M. Phytochemical analysis of Terminalia catappa stem using GC-MS/MS. *Res J Pharm Technol.* 2015; 8(9): 1281–3. doi: 10.5958/0974-360X.2015.00232.2
 48. Pandian RS, Noora AT. GC-MS analysis of phytochemical compounds present in the leaves of Citrus medica. L. *Res J Pharm Technol.* 2019; 12(4): 1823–6. doi: 10.5958/0974-360X.2019.00304.4
 49. Azmi L, Gupta SS, Shukla I, Kant P, Sidhu OP, Rao C V. Effect of squalene in surgically induced gastro-oesophageal reflux disease on rats. *Res J Pharmacol Pharmacodyn.* 2017; 9(1): 1. doi: 10.5958/2321-5836.2017.00001.5
 50. Sanjeev PA, Doshi GM. Chemical constituents from polyalthia longifolia seeds extract by gas chromatography-mass spectroscopy (GC-MS) studies. *Res J Pharm Technol.* 2018; 11(6): 2489–92. doi: 10.5958/0974-360X.2018.00459.6
 51. Kim SJ, Nara E, Kobayashi H, Terao J, Nagao A. Formation of cleavage products by autooxidation of lycopene. *Lipids.* 2001; 36(2):191. doi: 10.1007/s11745-001-0706-8
 52. Hazra A, Dutta M, Dutta R, Bhattacharya E, Bose R, Biswas SM. Squalene synthase in plants – Functional intricacy and evolutionary divergence while retaining a core catalytic structure. *Plant Gene.* 2023; 33: 100403. doi: 10.1016/j.plgene.2023.100403
 53. Gohil N, Bhattacharjee G, Khambhati K, Braddick D, Singh V. Engineering strategies in microorganisms for the enhanced production of squalene: Advances, challenges and opportunities. *Front Bioeng Biotechnol.* 2019; 7(MAR): 1–24. doi: 10.3389/fbioe.2019.00050
 54. Peng W, Li D, Zhang M, Ge S, Mo B, Li Shasha, Ohkoshi M. Characteristics of antibacterial molecular activities in poplar wood extractives. *Saudi J Biol Sci.* 2017; 24(2):399–404. doi: 10.1016/j.sjbs.2015.10.026
 55. Paul S, Mallick A, Majumdar T. Computational study on the ion interaction of ellipticine: A theoretical approach toward selecting the appropriate anion. *Chem Phys Lett.* 2015 Aug;634:29–36. doi: 10.1016/j.cplett.2015.05.051
 56. Vann KR, Ergün Y, Zencir S, Oncuoglu S, Osheroff N, Topcu Z. Inhibition of human DNA topoisomerase IIa by two novel ellipticine derivatives. *Bioorg Med Chem Lett.* 2016; 26(7): 1809–12. doi: 10.1016/j.bmcl.2016.02.034
 57. Barreto JN, McCullough KB, Ice LL, Smith JA. Antineoplastic agents and the associated myelosuppressive effects: A review. *J Pharm Pract.* 2014; 27(5): 440–6. doi: 10.1177/089719001454
 58. Borah N, Gogoi D, Ghosh NN, Tamuly C. GA-AuNP@Tollens' complex as a highly sensitive plasmonic nanosensor for detection of formaldehyde and benzaldehyde in preserved food products. *Food Chem.* 2023; 399: 133975. doi: 10.1016/j.foodchem.2022.133975
 59. Ogrande JL, Dobratz CJ, Brown E, Liang J, Conn GS, Shelton FJ., With J. Benzaldehyde. *Kirk-Othmer Encycl Chem Technol.* 2000; doi: 10.1002/0471238961.0205142615160718.a01
 60. Hoseini SM, Gharavi B, Taberi MA, Hoseinifar SH, Van Doan H. Effects of dietary phytol supplementation on growth performance, immunological parameters, antioxidant and stress responses to ammonia exposure in common carp, *Cyprinus carpio* (Linnaeus, 1758). *Aquaculture.* 2021; 545: 737151. doi: 10.1016/j.aquaculture.2021.737151
 61. Usman MA, Usman FI, Abubakar MS, Salman AA, Adamu A, Ibrahim MA. Phytol suppresses parasitemia and ameliorates anaemia and oxidative brain damage in mice infected with *Plasmodium berghei*. *Exp Parasitol.* 2021; 224: 108097. doi: 10.1016/j.exppara.2021.108097
 62. Guimarães A, Venâncio A. The Potential of Fatty Acids and Their Derivatives as Antifungal Agents: A Review. *Toxins* 2022; 14: 188. 2022 Mar;14(3):188.
 63. Park JS, Rhee SD, Kang NS, Jung WH, Kim HY, Kim JY, Kang SK, Cheon HG, Ahn JH, Kim KY. Anti-diabetic and anti-adipogenic effects of a novel selective 11 β -hydroxysteroid dehydrogenase type 1 inhibitor, 2-(3-benzoyl)-4-hydroxy-1,1-dioxo-2H-1,2-benzothiazine-2-yl-1-phenylethanone (KR-66344). *Biochem Pharmacol.* 2011; 81(8): 1028–35.
 64. Jiang C, Jiang Z, Zhu S, Amulraj J, Deenadayalan VK, Jacob J.A., Qian J. Biosynthesis of silver nanoparticles and the identification of possible reductants for the assessment of in vitro cytotoxic and in vivo antitumor effects. *J Drug Deliv Sci Technol.* 2021; 63: 102444. doi: 10.1016/j.jddst.2021.102444
 65. Murugan S, Nehru J, Arputharaj DS, Catherine Paul A, Gunasekaran P, Dege N, Berrin E, Balasubrahmani K, Savaridasson JK, Al-sehemi AG, Rajakannan V, Hemamalini M. Synthesis of 3-Methoxy-6- [(2, 4, 6-trimethyl-phenylamino)-methyl]-phenol Schiff base characterized by spectral, in-silico and in-vitro studies. *Heliyon.* 2022; 8(8): e10070. doi: 10.1016/j.heliyon.2022.e10070
 66. Raman Ibrahim NBB, Puchooa D, Govinden-Soulange J, Facknath S. Chemical profiling and biological activity of Cassia abbreviata Oliv. *South African J Bot.* 2022; 146: 325–39. doi: 10.1016/j.sajb.2021.11.004
 67. Sofidiya MO, Alokun AM, Fageyinbo MS, Akindele AJ. Central nervous system depressant activity of ethanol extract of *Motandra guineensis* (Thonn) AD. aerial parts in mice. *Phytomedicine Plus.* 2022; 2(1): 100186. doi: 10.1016/j.phyplu.2021.100186
 68. Umar HI, Awonyemi IO, Abegunde SM, Igbe FO, Siraj B. In Silico Molecular Docking of Bioactive Molecules Isolated from *Raphia taedigera* Seed Oil as Potential Anti-cancer Agents Targeting Vascular Endothelial Growth Factor Receptor-2. *Chem Africa.* 2021; 4(1): 161–74. doi: 10.1007/s42250-020-00206-8
 69. N-(1,1,3,3-tetramethylbutyl)-1-naphthalenamine | Sigma-Aldrich.
 70. Staniszevska M, Bondaryk M, Ochal Z. Role of Virulence Determinants in *Candida albicans*' Resistance to Novel 2-bromo-2-chloro-2-(4-chlorophenylsulfonyl)-1-phenylethanone. *J Fungi.* 2017; 3: 32. doi: 10.3390/jof3030032
 71. Dionisio KL, Phillips K, Price PS, Grulke CM, Williams A, Biryol D, Hong T, Isaacs KK. Data Descriptor: The Chemical and Products Database, a resource for exposure-relevant data on chemicals in consumer products. *Sci Data.* 2018;5. doi: 10.1038/sdata.2018.125
 72. Araújo GF, Carneiro LMV, Junior APF, Vieira LC, Bandeira PN, Lemos TLG, Viana GS d. B. A possible mechanism for anxiolytic and antidepressant effects of alpha- and beta-amyryn from *Protium heptaphyllum* (Aubl.) March. *Pharmacol Biochem Behav.* 2006; 85(4): 827–34. doi: 10.1016/j.pbb.2006.11.019
 73. Khajuria V, Gupta S, Sharma N, Kumar A, Lone NA, Paul S, Meena SR, Satti NK, Kumar C, Sharma A, Verma MK, Ahmed Z. Anti-inflammatory potential of hentricontane in LPS stimulated RAW 264.7 cells and mice model. *Biomed Pharmacother.* 2017; 92: 175–86. doi: 10.1016/j.biopha.2017.05.063
 74. Vélez C, Khayet M, Ortiz De Zárate JM. Temperature-dependent thermal properties of solid/liquid phase change even-numbered n-alkanes: n-Hexadecane, n-octadecane and n-eicosane. *Appl Energy.* 2015 Apr;143:383–94. doi: 10.1016/j.apenergy.2015.01.054
 75. Das A, Mishra M, Jaison JP, Sebastian JK. Phenolic composition and antioxidant potential of *Cosmosigma cordatum*. *Med Plants.* 2022; 14(4): 597–603. doi: 10.5958/0975-6892.2022.00065.X
 76. Fernando IPS, Sanjeeva KKA, Ann YS, Ko C ik, Lee SH, Lee WW, Jeon YJ. Apoptotic and antiproliferative effects of Stigmast-5-en-3-ol from *Dendronephthya gigantea* on human leukemia HL-60 and human breast cancer MCF-7 cells. *Toxicol Vit.* 2018; 52: 297–305. doi: 10.1016/j.tiv.2018.07.007
 77. Laparra JM, Alfonso-García A, Alegría A, Barberá R, Cilla A. 7keto-stigmastrol and 7keto-cholesterol induce differential proteome changes to intestinal epithelial (Caco-2) cells. *Food Chem Toxicol.* 2015 Oct;84:29–36. doi: 10.1016/j.fct.2015.06.021
 78. Bae H, Song G, Lim W. Stigmastrol causes ovarian cancer cell apoptosis by inducing endoplasmic reticulum and mitochondrial dysfunction. *Pharmaceutics.* 2020; 12(6). doi: 10.3390/pharmaceutics12060488
 79. Brigelius-Flohé R, Traber MG. Vitamin E: function and metabolism. *FASEB J.* 1999; 13(10): 1145–55. doi: 10.1096/fasebj.13.10.1145
 80. Behfar AA, Shushizadeh MR, Far MH, Shoar TS, Farasat M, Ghotrami ER. Gas chromatography-mass evaluation of terpenoids from Persian Gulf *Padina tetrastratica* sp. *Asian J Pharm.* 2018; 12(4): S115–9.

Phytoconstituents profiling of *Selaginella willdenowii* (Desv.) Baker and Pharmacological Potential

ORIGINALITY REPORT

20%

SIMILARITY INDEX

17%

INTERNET SOURCES

12%

PUBLICATIONS

%

STUDENT PAPERS

PRIMARY SOURCES

- | | | |
|---|--|----|
| 1 | www.mdpi.com
Internet Source | 2% |
| 2 | RC Cambie. "Ring A Modifications of Podocarpic Acid: Towards the Synthesis of Quassinoids", <i>Australian Journal of Chemistry</i> , 1991
Publication | 2% |
| 3 | pqr.pitt.edu
Internet Source | 1% |
| 4 | www.jazindia.com
Internet Source | 1% |
| 5 | www.lookchem.cn
Internet Source | 1% |
| 6 | Suaad Nasrin, Mohammad Nazmul Islam, Mohammed Abu Tayab, Mst. Samima Nasrin et al. "Chemical profiles and pharmacological insights of <i>Anisomeles indica</i> Kuntze: An experimental chemico-biological interaction", <i>Biomedicine & Pharmacotherapy</i> , 2022
Publication | 1% |

7	www.ijpsonline.com Internet Source	1 %
8	repositorio.ufsc.br Internet Source	1 %
9	www.knowitall.com Internet Source	1 %
10	hdl.handle.net Internet Source	1 %
11	www.teknolabjournal.com Internet Source	1 %
12	www.isca.in Internet Source	1 %
13	coek.info Internet Source	<1 %
14	ojshostng.com Internet Source	<1 %
15	rjas.org Internet Source	<1 %
16	www.chemspider.com Internet Source	<1 %
17	Njoku Ugochi Olivia, Umeh Chinenyenwa Goodness, Ogugofor Martins Obinna. "Phytochemical profiling and GC-MS analysis of aqueous methanol fraction of Hibiscus	<1 %

asper leaves", Future Journal of Pharmaceutical Sciences, 2021

Publication

18

www.tropicalplantresearch.com

Internet Source

<1 %

19

www.iptsalipur.org

Internet Source

<1 %

20

www.thuvientailieu.vn

Internet Source

<1 %

21

Susilo, M Setyaningsih. " Analysis of genetic diversity and genome relationships of four eggplant species (L) using RAPD markers ", Journal of Physics: Conference Series, 2018

Publication

<1 %

22

ictromi.usu.ac.id

Internet Source

<1 %

23

mdpi-res.com

Internet Source

<1 %

24

May Poh Yik GOH, Ajmal Faiz KAMALUDDIN, Terence Jit Loong TAN, Hartini YASIN, Hussein TAHA, Abdalla JAMA, Norhayati AHMAD. "An evaluation of the phytochemical composition, antioxidant and cytotoxicity of the leaves of Litsea elliptica Blume - an ethnomedicinal plant from Brunei Darussalam", Saudi Journal of Biological Sciences, 2021

Publication

<1 %

25	assets.researchsquare.com Internet Source	<1 %
26	nlist.inflibnet.ac.in Internet Source	<1 %
27	repositorio.ufpe.br Internet Source	<1 %
28	www.pherobase.com Internet Source	<1 %
29	journal.uinsgd.ac.id Internet Source	<1 %
30	jurnal.ugm.ac.id Internet Source	<1 %
31	nopr.niscair.res.in Internet Source	<1 %
32	www.phytojournal.com Internet Source	<1 %
33	I.P. Shanura Fernando, K.K. Asanka Sanjeewa, Yong-Seok Ann, Chang-ik Ko, Seung-Hong Lee, Won Woo Lee, You-Jin Jeon. "Apoptotic and antiproliferative effects of Stigmast-5-en-3-ol from Dendronephthya gigantea on human leukemia HL-60 and human breast cancer MCF-7 cells", Toxicology in Vitro, 2018 Publication	<1 %
34	cms.turkjps.org	

Internet Source

<1 %

35

ijlpr.com

Internet Source

<1 %

36

www.iosrjournals.org

Internet Source

<1 %

37

www.researchgate.net

Internet Source

<1 %

38

Natural Products, 2013.

Publication

<1 %

39

"Natural Product Experiments in Drug
Discovery", Springer Science and Business
Media LLC, 2023

Publication

<1 %

Exclude quotes On

Exclude matches Off

Exclude bibliography On