

# Numlil - CANCER IMMUNOTHERAPY AND FLOW CYTOMETRY IN IMMUNOTHERAPY MONITORING

*by* Numlil Uploader Lutfan Zulwaqar

---

**Submission date:** 03-Oct-2019 05:19PM (UTC+0700)

**Submission ID:** 1185212059

**File name:** rev\_1\_ARTIKEL-Numlil-English-final.docx (191.78K)

**Word count:** 3510

**Character count:** 20557

## **CANCER IMMUNOTHERAPY AND FLOW CYTOMETRY IN IMMUNOTHERAPY MONITORING**

**Numlil Khaira Rusdi<sup>1,2</sup>, Jeanne Adiwinata Pawitan<sup>3,4,5</sup>**

1. Doctoral Program for Biomedical Sciences, Faculty of Medicine, Universitas Indonesia, Jakarta, Indonesia

2. Faculty of Pharmacy and Science, University Muhammadiyah Prof. DR. Hamka, Jakarta, Indonesia

3. Department of Histology, Faculty of Medicine, Universitas Indonesia, Jakarta, Indonesia

4. Stem Cell Medical Technology Integrated Service Unit, Dr. Cipto Mangunkusumo General Hospital/Faculty of Medicine Universitas Indonesia, Indonesia

5. Stem Cell and Tissue Engineering Research Center, Indonesia Medical Education and Research Institute (IMERI), Faculty of Medicine Universitas Indonesia, Indonesia

### **Corresponding author:**

Jeanne Adiwinata Pawitan

Department of Histology, Faculty of Medicine Universitas Indonesia, Jakarta, Indonesia

Email: [jeanneadiwip@gmail.com](mailto:jeanneadiwip@gmail.com)

## ABSTRACT

Immunotherapy for cancer treatment continues to be developed and various strategies have been carried out including bioengineering. This endeavour requires development of technology, and efforts to find specific and sensitive tools to monitor immune responses during and after therapy. The purpose of this mini-review was to discuss cancer immunotherapy using T cell and immune checkpoint blockade therapy, as well as immunotherapy monitoring methods using flow cytometry (FCM). Bioengineering of T lymphocytes for immunotherapy and immune checkpoint blockades can be combined with nanoparticles as drug delivery carrier against cancer to increase drug distribution to tumor cells, as well as T cell stimulation regulation to reduce autoimmune effects. In addition, T cell engineering can also prevent Host versus Graft alloreactivity in chimeric antigen receptor (CAR) T cell administration. FCM is a monitoring method that is widely used in pre-clinical and clinical cancer immunotherapy studies.

**Keywords:** Bioengineering, Cancer immunotherapy, T cells, Immune checkpoint, Flow cytometry

## INTRODUCTION

Immunotherapy is a method that is continuously developed in the treatment of cancer.<sup>1,2</sup> Immunotherapy has a great potential to treat and prevent cancer recurrence by activating the immune system to recognize and kill cancer cells.<sup>3</sup> Clinical studies using chimeric antigen receptor (CAR) T cells and immune checkpoint blockade are examples of the success in cancer immunotherapy.<sup>4,5</sup> Adoptive Cell Therapy (ACT) using tumor infiltration lymphocytes (TIL) has also been used successfully in treating patients with metastatic melanoma.<sup>6</sup>

Various immunotherapy strategies continue to be developed including cancer and dendritic cell vaccines, T cell engineering, and immune checkpoint blocking.<sup>1,3</sup> Although it is believed that the immune system can provide an immune response to the growth of malignant tumors, in fact, many malignant tumors still grow in immunocompetent individuals.<sup>7,8</sup> Therefore, research in immunotherapy also needs to be conducted to understand how and why certain cancers fail to respond to immunotherapy, and also to develop appropriate immunotherapy strategies.<sup>3</sup>

This endeavour requires development of technology, including rapid methods to characterize the immune microenvironment of tumors, to find specific and sensitive tools to assess patient therapeutic outcomes, and to make it possible to monitor immune responses, tumor growth, and spread of tumors during and after therapy.<sup>3</sup> One method to overcome these problems is the use of immunoengineering in cancer therapy.<sup>9</sup> In addition, methods for immunotherapy monitoring are needed, and one of these methods that are widely used is flow cytometry.<sup>10,11</sup>

Flow cytometry (FCM) is a multi-parameter analytic test for the characterization of single cells and has been widely used in preclinical as well as in clinical cancer immunotherapy studies.<sup>12,13</sup> The aim of this mini-review was to discuss cancer immunotherapy using T cell and immune checkpoint blockade therapy, as well as immunotherapy monitoring methods using FCM.

## T CELL IMMUNOTHERAPY FOR CANCER

Engineering technology has been developed to improve cancer therapy using T cells. One of them is the use of nanocarriers as drug delivery vehicle (carrier) that are incorporated on the surface of T cells for cancer therapy. Nanocarriers are often referred to as nanoparticles, nanomaterials, or nanosomes. The use of nanoparticles as a drug delivery carrier in cancer therapy aims to increase the bioavailability and payload (distribution) of drugs, and to improve drug release to tumor cells, as well as to improve T cell function in infiltrating tumors and to minimize autoimmune toxicity.<sup>14,15</sup>

To avoid recognition by the immune system, cancer cells will activate Shp1/2 phosphatase in TIL to stimulate T cell inhibitory receptors such as PD-1, and CTLA-4. Moreover, Shp1/2 phosphatases migrate to the immunological synapse (IS), deactivate molecular signals of proliferation, and down-regulate T cell activation.<sup>14</sup> Stephan et al.<sup>14</sup> engineered T lymphocytes by conjugating a synthetic nanocarrier NSC-87877 to the surface of T lymphocytes. NSC-87877 is a phosphatase inhibitor, which inhibit Shp1/2 phosphatase. Therefore conjugation of

NSC-87877 synthetic nanocarrier on the surface of T lymphocytes can activate T cells, increase immunologic synapses, and facilitate molecular interaction between T cells and APCs. This interaction causes activation of tumor antigen recognition by APC cells, increases T cell (TIL) function in tumors, and minimizes autoimmune toxicity. Engineered T cell-synthetic nanocarrier NSC-87877 were injected to prostate cancer mice models through the tail vein, and the distribution was observed using bioluminescent imaging. This study proved the expansion of T cells in tumor sites resulting in an increase in the survival of experimental animals with prostate tumors. In addition, T cells conjugated with synthetic nanocarrier NSC-87877 could activate immunological synapses, and concentrate NSC-87877 in the contact zone of T-cell/tumor cell based on "demand", so that it could regulate T cell stimulation and reduce autoimmune reactions.<sup>14</sup>

Other studies also proved that T lymphocytes could be engineered as carriers of topoisomerase-1 SN-38 that were effectively transported to the cancer site.<sup>16</sup> Topoisomerase-1 SN-38 is a systemic chemotherapy, which is an active form of camptothecin derivative of Irinotecan. This drug has poor pharmacokinetics. To increase its biodistribution and effectiveness, researchers engineered polyclonal autologous T lymphocytes as carriers of topoisomerase 1 SN-38 in the form of controlled release lipid nanocapsules (NCs), namely SN-38 NC-T cells. As lymphocytes migrate throughout the tissue to look for antigens, polyclonal T cells, which express lymph node receptors but do not specifically recognize tumor cell antigens, can be used as an effective agent to deliver chemotherapy drugs for lymphoma. SN-38 NC-T cells were tested on Burkitt's lymphoma animal model, and their effectiveness and toxicity were observed. The results showed that SN-38 NC-T cell therapy significantly increased the efficacy of SN-38 without increasing the drug side effects.<sup>16</sup>

The application of T cell immunotherapy was also developed by Valton, et al.,<sup>17</sup> where engineering was performed to prevent host versus graft alloreactivity in the administration of T cell chimeric antigen receptor (CAR) therapy. CAR T-cell therapy was obtained by taking T cells' patients with leukapheresis procedures.<sup>18</sup> T cells taken were then engineered in the laboratory. The engineered T cell receptors could recognize specific antigens in cancer cells, which would destroy cancer cells specifically. This modified T cell was called the chimeric antigen receptor T-cell (CAR T-cell). T cells that had been modified were then reinfused into the patient's body.<sup>19</sup>

Giving infusion of CAR T-cell might cause alloreactivity in patients. To prevent this reaction, Valton, et al.,<sup>17</sup> engineered T cells by inactivating TCR $\alpha\beta$  and purine nucleotide analogs on the surface of T cells to prevent alloreactivity and to prevent damage of lymphocytes through host versus graft (HvG) reactions. T-cell engineering includes lentiviral transduction for CAR expression followed by inactivation "TCR $\alpha\beta$  within CAR-T cell genome", through inactivation of *deoxycytidine kinase* (dCK), and coelectroporation with mRNA that coded TCR constant region (TRAC) transcription activator-like effector nuclease (TALEN). DCK is responsible for TCR $\alpha\beta$  expression and toxicity of purine nucleotide analogs (PNA), which can damage the immune system. PNA becomes toxic after being metabolized by dCK. Therefore dCK inactivation prevents PNA metabolism and toxicity, thus protect the immune system. Inactivation of dCK was done by designing TALEN that targeted the second exon of dCK gene, and TALEN mRNA

was used to electroporate primary T cells. This approach was reported to be efficient in inactivating the TCR $\alpha$  gene and preventing the expression of TCR $\alpha\beta$ .<sup>17</sup>

## IMMUNE CHECKPOINT BLOCKADE THERAPY

Included in the immune checkpoint are cytotoxic T-lymphocyte-associated protein 4 (CTLA-4), programmed cell death protein 1 (PD-1 / PDL-1), Human B7-H3 (CD276) and Lymphocyte-activation gene-3 (LAG-3, CD223).<sup>20</sup> Immune checkpoint inhibitors that focus on CTLA-4 and PD-1 have been shown to have a potential of immunomodulators through their function as negative activators of T cells.<sup>5,21,22</sup> CTLA-4 plays an important role in reducing activation of memory T cells and naive T cells. In contrast, PD-1 is mainly involved in modulating T cell activity in peripheral tissues through its interaction with PD-L1 and PD-L2.<sup>5</sup>

The discovery of a negative regulator of this immune response is very important in the development of immune checkpoint inhibitors. This shifts the focus of research from developing therapies that target immune system activation against cancer to immune checkpoint inhibitors, which aim to mediate the destruction of tumor cells through coinhibitory signal removal that blocks antitumor T cell responses.<sup>5,23</sup>

One of the applications of immune checkpoint blocking therapy was the study by Zhang *et al.*<sup>24</sup>, who performed genetic engineering of platelets from megakaryocyte (MK) progenitor cells to express PD-1 protein. This study aimed to prevent postoperative tumor recurrence.<sup>24</sup> Genetically engineered platelets of MK cells that express PD-1/ PD-1 platelets and derivative microparticles accumulated in post-tumor surgery wounds, and restore CD8 + T cell state to lyse residual tumor cells. Furthermore, low-dose cyclophosphamide (CP) was inserted into PD-1 expression platelets to destroy regulatory T cells (Treg), as well as to increase CD8 + lymphocyte cells that were active, to directly prevent tumor recurrence.<sup>24</sup>

PD-1 expression platelets were produced from the L8057 cell line labeled with NHS-Cy5.5 in PBS. This labeled platelets were then infused into mice with 10% residual melanoma tumors after tumor removal. The study used 42 mice that were divided into 6 treatment groups, which were given IV PBS, free platelets, PD-1 platelets, cyclophosphamide, cyclophosphamide-loaded free platelets, or cyclophosphamide-loaded PD-1-expressing platelets. Tumor burden was monitored using bioluminescence. The parameters observed were the frequency of Treg (CD4 + Treg), CD8 + TIL, PD-L1, GzmB expression, and Ki67.

The results showed that the utilization of cyclophosphamide-loaded PD-1 expression platelets effectively disrupted PD-L1 immune blocking and reduced Treg, and significantly increased the frequency of CD8 + TIL in tumors, thereby reducing the rate of post-removal tumor recurrence. In addition, PD-1 expression platelets promoted the emergence of CD8 +, Ki67, and GzmB lymphocytes in the postremoval tumor microenvironment.

## FLOW CYTOMETRY IN MONITORING CANCER IMMUNOTHERAPY

The discovery of successful immunotherapy in cancer requires specific and sensitive tools to assess patient therapeutic outcomes and makes it possible to monitor immune response, tumor growth, and spread of tumors during and after therapy.<sup>3</sup> Immune system monitoring becomes increasingly relevant to measure the effects of treatment on T lymphocytes and to explain the mechanism of successful treatment.<sup>25</sup>

Flow cytometry (FCM) is a multi-parameter test for the characterization of single cells, and has been widely used in pre-clinical and clinical cancer immunotherapy studies.<sup>25</sup> FCM is a technique of measuring the number and characteristics of single cells that are flowing through a detector system, using a fluorochrome-labeled monoclonal antibody. Commonly used fluorochromes are fluorescein isothiocyanate (FITC), phycoerythrin (PE), peridinin-chlorophyll alpha complex (PerCP), allophycocyanin (APC), and propidium iodide (PI).<sup>26</sup>

4 The FCM application in monitoring cancer immune systems includes intracellular staining for effector cytokines, cytotoxicity evaluation, measurement of proliferation and evaluation of cells that regulate the immune system including regulatory T cells (T reg).<sup>25</sup> FCM allows quantization of several parameters in many individual cells that combines various phenotypic and functional markers and has the potential to provide information about immune responses in cancer patients.<sup>10,25</sup>

In FCM where the sample sources might come from peripheral blood, bone marrow, body fluids, urine, spleen cells or tissue, or cultured cell line, the cells should be processed into cell suspension. The cell suspension is mixed with fluorochrome labeled antibodies, and injected into a fluidic system, where the flow pushes the cells to pass through a central channel to line up as single cells as they approach the laser. Lasers as light sources will excite the cells in the stream and optical filters direct the resulting light signals to the appropriate detectors. The detector that detects the light scatter in forward direction shows the forward scatter (FSC) as the X-axis, and the detector that detects deflected light scatter shows the side scatter (SSC) as the Y-axis. FSC correlates with cell size, while the SSC is related to the complexity of the cells, such as nucleus size, types and complexity of cytoplasmic granules.

Interpretation of FCM results in the form of cell quantification and cell characteristics. The Scatter plot is a visualization of characteristics that is commonly used in FCM results. X-axis is as FCS and Y axis is as SSC. The X and Y axes can also be based on fluorochromes and surface markers on cells, for example: FITC-CD3 (CD3 surface markers labelled by FITC fluorochrome). Fluorochrome on the X and Y axes must be different. The results are translated into electronic signals and graphical displays. The signals are converted into digital numbers and shown on a histogram, 2-D or 3-D plot.

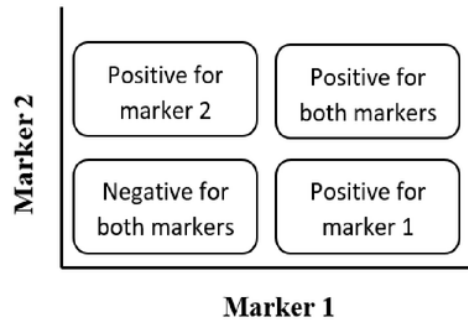


Figure 1. Interpretation of Flow Cytometry Result

In a 2D plot, if the cell plot is in the upper right quadrant, then the cell is positive for marker 1 and marker 2, whereas if it is in the lower left quadrant, then the cell is negative for both markers. If the plot is in the lower right quadrant, the cell is positive for marker 1, and if the cell plot is in the upper left, the cell is positive for marker 2 (Figure 1).

The results of the FCM examination of cell quantification is in the form of a cell percentage, while to get an absolute number should be done by counting cells in a hemacytometer. FCM requires a lot of fluorochrome-labeled antibodies (according to parameters), which is a disadvantage. However it has some advantages, i.e.: (a) FCM has a high level of efficiency and sensitivity, and can obtain information simultaneously about various cellular processes, including expression of cell surface markers, intracellular cytokines, protein signals, and cell cycles, (b) the process is fast and enable quantitative measurements of many parameters simultaneously, (c) High specificity (can detect cells that are small in number) and (d) isolation of cells with high cell purity.<sup>13</sup>



**Table 1. Cancer immunotherapy using T cells and blockade immune checkpoints**

No	Immuno therapy	Purpose of engineering	Type of engineering	Type of Study	Analytical Instrument
1	T cells Therapy	<ul style="list-style-type: none"> <li>▪ Using nanoparticles as a Drug Delivery vehicle (carrier) for cancer therapy to increase the payload (distribution) of drugs to tumor cells.</li> <li>▪ The use of T cells as the target delivery of the nanoparticle topoisomerase I-SN-38 drug</li> <li>▪ Preventing Host versus Graft Alloreactivity in the administration of T cell infusion of chimeric antigen receptor (CAR) with T cell tissue engineering models</li> </ul>	<ul style="list-style-type: none"> <li>▪ Surface engineering of T lymphocytes with synthetic nanocarrier NSC-87877</li> <li>▪ Engineering polyclonal autologous T cells conjugated with SN-38 topoisomerase I nanocapsules</li> <li>▪ Engineering T cells by inactivating TCRαβ and purine nucleotide analogs on the surface of T cells, that can prevent alloreactivity and to prevent damage of lymphocyte cells through host versus graft reaction (HvG)</li> </ul>	<p><i>Animal study</i></p> <p><i>Animal study</i></p> <p><i>Clinical Study</i></p>	<ul style="list-style-type: none"> <li>▪ Flowcytometry</li> <li>▪ Confocal and videomicroscopy</li> <li>▪ Bioluminescent imaging</li> <li>▪ Flowcytometry</li> <li>▪ IVIS imaging</li> <li>▪ Bioluminescent imaging</li> <li>▪ Flowcytometry</li> <li>▪ Spectrophotometry (ELISA)</li> <li>▪ Nested PCR</li> <li>▪ Magnetic separation</li> </ul>
2	<i>Immune Blockade Therapy</i>	<ul style="list-style-type: none"> <li>▪ Development of a drug delivery system that utilizes PD-1 expression platelets for the improvement of postoperative cancer immunotherapy.</li> </ul>	<ul style="list-style-type: none"> <li>▪ PD-1 expression platelets was produced from the L8057 cell line labeled NHS-Cy5.5. In PD-1 expression platelets given a loading dose of cyclophosphamide.</li> </ul>	<p><i>Animal study</i></p>	<ul style="list-style-type: none"> <li>▪ Flowcytometry</li> <li>▪ Western Blot</li> <li>▪ Confocal microscopy, CSEM, TEM images</li> <li>▪ Xenogen IVIS Lumina imaging</li> </ul>

## CONCLUSION

Engineering of T lymphocytes for cancer immunotherapy and immune checkpoint blockade can be combined with nanoparticles as a drug delivery carrier to increase drug distribution to tumor cells and reduce autoimmunity. In addition, T cell engineering was proven in preventing Host versus Graft alloreactivity in the administration of CAR T cells. FCM is a monitoring method widely used in cancer immunology in pre-clinical and clinical cancer immunotherapy studies.

## Competing of interest

The authors declare that they have no competing interest.

## REFERENCES

1. Zhang H, Chen J. Current status and future directions of cancer immunotherapy. *J Cancer*. 2018 Apr 19;9(10):1773–81. doi: 10.7150/jca.24577
2. Yu LY, Tang J, Zhang CM, Zeng WJ, Yan H, Li MP, Chen XP. New Immunotherapy Strategies in Breast Cancer. *Int J Environ Res Public Health*. 2017 Jan 12; 14(1). pii: E68. doi: 10.3390/ijerph14010068
3. Jeanbart L, Swartz MA. Engineering opportunities in cancer immunotherapy. *PNAS 100th Anniversary*. 2015 Nov;112(47):14467–72.
4. Hou B, Tang Y, Li W, Zeng Q, Chang D. Efficiency of CAR-T Therapy for Treatment of Solid Tumor in Clinical Trials : A Meta-Analysis. *Dis Markers*. 2019 Feb 11;2019:3425291 (11 pages). doi: 10.1155/2019/3425291.
5. Webb ES, Liu P, Baleeiro R, Lemoine NR, Yuan M, Wang Y. Immune checkpoint inhibitors in cancer therapy. *J Biomed Res*. 2018 Sep 29;32(5):317–26. doi: 10.7555/JBR.31.20160168.
6. Rohaan MW, van den Berg JH, Kvistborg P, Haanen JBAG. Adoptive transfer of tumor-infiltrating lymphocytes in melanoma : a viable treatment option. *J Immunother Cancer*. 2018 Oct 3;6(1):102. doi: 10.1186/s40425-018-0391-1
7. Sambhi M, Bagheri L, Szewczuk MR. Current Challenges in Cancer Immunotherapy : Multimodal Approaches to Improve Efficacy and Patient Response Rates. *J Oncol*. 2019 Feb 28;2019:4508794 (12 pages). doi: 10.1155/2019/4508794.
8. Zhu J, Powis de Tenbossche CG, Cané S, Colau D, van Baren N, Lurquin C, Schmitt-Verhulst AM, Liljeström P, Uyttenhove C, Van den Eynde BJ. Resistance to cancer immunotherapy mediated by apoptosis of tumor-infiltrating lymphocytes. *Nat Commun*. 2017 Nov 10;8(1):1404. doi:

- 10.1038/s41467-017-00784-1.
9. Lim WA, June CH. The Principles of Engineering Immune Cells to Treat Cancer. *Cell*. 2017 Feb 9;168(4):724-40. doi: 10.1016/j.cell.2017.01.016.
  10. Danova M, Torchio M, Comolli G, Sbrana A, Antonuzzo A, Mazzini G. The role of automated cytometry in the new era of cancer immunotherapy. *Mol Clin Oncol*. 2018 Oct;9(4):355-61. doi: 10.3892/mco.2018.1701.
  11. Chargin A, Morgan R, Sundram U, Shults K, Tsay EL, Ratti N, Patterson BK. Quantification of PD - L1 and PD - 1 expression on tumor and immune cells in non - small cell lung cancer ( NSCLC ) using non - enzymatic tissue dissociation and flow cytometry. *Cancer Immunol Immunother*. 2016 Nov;65(11):1317–23. doi 10.1007/s00262-016-1889-3.
  12. Gascue A, Merino J, Paiva B. Flow Cytometry. *Hematol Oncol Clin North Am*. 2018 Oct;32(5):765-75. doi: 10.1016/j.hoc.2018.05.004.
  13. Jahan-Tigh RR, Ryan C, Obermoser G, Schwarzenberger K. Flow Cytometry. *J Invest Dermatol*. 2012 Oct;132(10):1-6. doi: 10.1038/jid.2012.282.
  14. Stephan MT, Stephan SB, Bak P, Chen J, Irvine DJ. Synapse-directed delivery of immunomodulators using T-cell conjugated nanoparticles. *Biomaterials*. 2012 Aug;33(23):5776–87. Doi: 10.1016/j.biomaterials.2012.04.029.
  15. Artini IG. Peranan Nanopartikel Dalam Penatalaksanaan Kanker di Era Targeting Therapy. *Indones J Cancer*. 2013;7(3):111–7.
  16. Huang B, Abraham WD, Zheng Y, Bustamante López SC, Luo SS, Irvine DJ. Active targeting of chemotherapy to disseminated tumors using nanoparticle-carrying T cells. *Sci Transl Med*. 2015 Jun 10;7(291):1-12. 291ra94. doi: 10.1126/scitranslmed.aaa5447.
  17. Valton J, Guyot V, Marechal A, Filhol JM, Juillerat A, Duclert A, Duchateau P, Poirot L. A Multidrug-resistant Engineered CAR T Cell for Allogeneic Combination Immunotherapy. *Mol Ther*. 2015 Sep;23(9):1507–18. doi: 10.1038/mt.2015.104.
  18. Juliano L, Eastwood G, Berard T, Mathew AJ. The Importance of Collection, Processing and Biopreservation Best Practices in Determining CAR-T Starting Material Quality. *Cell Gene Ther. Insights* 2018;4(4): 327–36. doi: 10.18609/cgti.2018.032
  19. Wang X, Rivière I. Clinical manufacturing of CAR T cells : foundation of a promising therapy. *Mol Ther Oncolytics*. 2016 Jun 15;3:1–7. 16015. doi: 10.1038/mto.2016.15.
  20. Galetto R, Lebuhotel C, Poirot L, Gouble A, Toribio ML, Smith J, Scharenberg A. Pre-TCR  $\alpha$  supports CD3-dependent reactivation and expansion of TCR  $\alpha$  -deficient primary human T-cells. *Mol Ther Methods Clin Dev*. 2014 Jun 11;1:1–9. 14021. doi: 10.1038/mtm.2014.21.

21. Park JA, Cheung NV. Limitations and opportunities for immune checkpoint inhibitors in pediatric malignancies. *Cancer Treat Rev.* 2017 Jul;58:22–33. doi: 10.1016/j.ctrv.2017.05.006.
22. Zhou Q, Munger ME, Highfill SL, Tolar J, Weigel BJ, Riddle M, Sharpe AH, Valleria DA, Azuma M, Levine BL, June CH, Murphy WJ, Munn DH, Blazar BR. Program death-1 signaling and regulatory T cells collaborate to resist the function of adoptively transferred cytotoxic T lymphocytes in advanced acute myeloid leukemia. *Blood J.* 2010 Oct 7;116(14):2484–93. doi: 10.1182/blood-2010-03-275446.
23. Dai F, Zhang F, Sun D, Zhang ZH, Dong SW, Xu JZ. CTLA4 enhances the osteogenic differentiation of allogeneic human mesenchymal stem cells in a model of immune activation. *Braz J Med Biol Res.* 2015 Jul;48(7):629–36. doi: 10.1590/1414-431X20154209.
24. Li Y, Li F, Jiang F, Lv X, Zhang R, Lu A, Zhang G. A Mini-Review for Cancer Immunotherapy : Molecular Understanding of PD-1/PD-L1 Pathway & Translational Blockade of Immune Checkpoints. *Int J Mol Sci.* 2016 Jul 18;17(7):1–22. pii: E1151. doi: 10.3390/ijms17071151.
25. Zhang X, Wang J, Chen Z, Hu Q, Wang C, Yan J, Dotti G, Huang P, Gu Z. Engineering PD-1-Presenting Platelets for Cancer Immunotherapy. *Nano Lett.* 2018 Sep 12;18(9):5716–25. doi: 10.1021/acs.nanolett.8b02321.
26. White S, Laske K, Welters MJ, Bidmon N, van der Burg SH, Britten CM, Enzor J, Staats J, Weinhold KJ, Gouttefangeas C, Chan C. Managing Multi-center Flow Cytometry Data for Immune Monitoring. *Cancer Inform.* 2015 Jun 10;13(Suppl 7):111-22. doi: 10.4137/CIN.S16346.
27. Adan A, Alizada G, Kiraz Y, Baran Y, Nalbant A. Flow cytometry : basic principles and applications. *Crit Rev Biotechnol.* 2017 Mar;37(2):163-176. doi: 10.3109/07388551.2015.1128876.

# Numlil - CANCER IMMUNOTHERAPY AND FLOW CYTOMETRY IN IMMUNOTHERAPY MONITORING

## ORIGINALITY REPORT

14%

SIMILARITY INDEX

12%

INTERNET SOURCES

13%

PUBLICATIONS

4%

STUDENT PAPERS

## PRIMARY SOURCES

1	<a href="http://journal.hep.com.cn">journal.hep.com.cn</a> Internet Source	3%
2	Xudong Zhang, Jinqiang Wang, Zhaowei Chen, Quanyin Hu, Chao Wang, Junjie Yan, Gianpietro Dotti, Peng Huang, Zhen Gu. "Engineering PD-1-Presenting Platelets for Cancer Immunotherapy", Nano Letters, 2018 Publication	2%
3	<a href="http://pubs.acs.org">pubs.acs.org</a> Internet Source	1%
4	Scott White, Karoline Laske, Marij J.P. Welters, Nicole Bidmon et al. "Managing Multi-center Flow Cytometry Data for Immune Monitoring", Cancer Informatics, 2015 Publication	1%
5	<a href="http://www.pnas.org">www.pnas.org</a> Internet Source	1%
6	Bonnie Huang, Wuhbet D. Abraham, Yiran Zheng, Sandra C. Bustamante López,	1%

Samantha S. Luo, Darrell J. Irvine. "Active targeting of chemotherapy to disseminated tumors using nanoparticle-carrying T cells", Science Translational Medicine, 2015

Publication

---

7	<a href="http://tessera.spandidos-publications.com">tessera.spandidos-publications.com</a> Internet Source	1%
8	<a href="http://biotechprimer.com">biotechprimer.com</a> Internet Source	1%
9	Submitted to University of Aberdeen Student Paper	1%
10	<a href="http://link.springer.com">link.springer.com</a> Internet Source	1%
11	<a href="http://orca.cf.ac.uk">orca.cf.ac.uk</a> Internet Source	<1%
12	<a href="http://www.ncbi.nlm.nih.gov">www.ncbi.nlm.nih.gov</a> Internet Source	<1%
13	<a href="http://pdfs.semanticscholar.org">pdfs.semanticscholar.org</a> Internet Source	<1%

---

Exclude quotes      On

Exclude matches      < 10 words

Exclude bibliography      On